



Reactions to honeybee stings: an allergic prospective

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Purpose of review

The purpose of this article is to provide a brief overview of the events involved in honeybee allergy and to concisely update the reader on progress toward knowledge of honeybee venom (HBV), strides in solving diagnostic difficulties, and advancements in improving safety and efficacy of HBV immunotherapy.

Recent findings

It is well known that honeybee allergy is unique in venom allergen and protein composition, diagnostic challenges, and immunotherapy safety and efficacy. Many new honeybee allergens have been recognized. Advances in testing, evaluation, and extract manipulation methods, many using recombinant technology, have allowed a greater ability to help with honeybee allergy diagnosis and resultant improvement in immunotherapy safety and evaluation of immunotherapy efficacy.

Summary

In an effort to address many honeybee allergy concerns, specific advances have been recently made. Some recently characterized honeybee allergens appear to be major contributors to honeybee allergy. In the setting of double-positivity, cross-reacting carbohydrate determinants and other cross-reacting components in HBV have made diagnosis of honeybee allergy challenging. Recombinant technology, including component-resolved diagnostics, and other evolving testing methods should help clarify double-positivity, if not now, in the very near future. Purified HBV and possibly depot formulations for immunotherapy appear to make it more well tolerated. Recombinant methods may help with evaluation of immunotherapy's safety and efficacy.

Keywords

component resolved diagnostics, cross-reacting carbohydrate determinant, honeybee allergy, recombinant venom allergen, venom immunotherapy

INTRODUCTION

To the practicing allergist, venom allergy is important because of the life-protecting implications of recognition and proper therapy. More than other flying Hymenoptera, honeybee allergy presents unique challenges. Diagnosis of true honeybee sensitization, especially in the face of double-positivity, and the efficacy and safety of honeybee venom immunotherapy (VIT) are often problematic and remain the focus of ongoing research. After reviewing honeybee sting reactions, this review summarizes recent findings in allergenic honeybee venom (HBV) components, progress to solve challenges in diagnosing honeybee sensitization and hence true allergy, and strides in making honeybee VIT more well tolerated and efficacious.

HONEYBEE STINGS

Reactions to honeybee stings range from small local reactions to large local reactions to anaphylaxis

and even death. Honeybee sting reactions may even be toxic, particularly mass envenomations, and cause rhabdomyolysis, hemolysis, thrombocytopenia, acute renal failure, hepatitis, mental status changes, and cardiac arrest [1,2]. Though solely honeybee data are lacking, stings seem to occur with a prevalence of about 30–40% among the adult population with a systemic reaction rate of 0.5–2% [3–5]. Death rates for honeybee stings are not well established, but one study [6] in the United States, likely underreported,

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Curr Opin Allergy Clin Immunol 2013, 13:365–371

DOI:10.1097/ACI.0b013e3283625144

KEY POINTS

- Honeybee allergy presents unique challenges in that its venom is more complex, it is more often the cause of double-positivity, and its venom immunotherapy is less well tolerated and less efficacious than other Hymenoptera.
- Three new HBV allergens appear to be significant contributors to honeybee allergy, one of which may be a new major allergen.
- Component-resolved diagnostics, using recombinant technology, may be useful in resolving double-positivity, though it is not ready to replace skin testing as the primary tool for evaluation.
- Tools for evaluating venom immunotherapy efficacy and safety have shown promise, and depot injection and purified extract injection are likely more well tolerated and equally effective compared with standard immunotherapy extracts.
- The basophil activation test may aid in both resolving double-positivity and in evaluating immunotherapy efficacy; however, it remains a difficult test to standardize and operate.

showed half of the 40 deaths per year were due to honeybees.

Eliciting stings

We tend to excite honeybees when we disturb their foraging behavior, typically through stepping on them or agitating them through abrupt, proximal movements (such as swatting or gardening) [7]. Fortunately, honeybees are docile, so stings are rare relative to encounters. The aggressiveness of honeybees depends on a number of factors to include air temperature, humidity, time of day, specific season, physical threat, honeybee species (Africanized honeybee being more aggressive), and perhaps odor [8–10]. Contrary to common conception, ‘looking like a flower’ shows little risk for a honeybee sting [7].

Honeybee sting mechanics

The honeybee stinging apparatus and mechanics have been described earlier [11]. Unique to the honeybee are multibarbed lancets that are prominent and that facilitate autotomy, making a sting more likely from a honeybee when the stinger is found at the sting site. Approximately, 50–140 µg of HBV on average is reported from a sting, with volumes as high as 300 µg [12,13]. It is important to note that honeybee sting volumes can vary widely,

as this may explain, at least in part, variable reactions during field stings and sting challenges.

Various routes of sensitization and reaction

Various routes of HBV sensitization and reaction have been reported recently, aside from the obvious sting and previously reported royal jelly, propolis, and other components of the hive [14,15]. A child experienced a systemic reaction after secondary mucocutaneous exposure to HBV on a glass from which he drank, which his father, a hobbyist beekeeper, had handled [16]. In a recent study [17] from Korea, it was noted that the only source of sensitization for 10% of their honeybee-allergic patients was from acupuncture with HBV extract, a common practice for pain relief in Oriental medicine. The important lesson is that sensitization and reaction can come through nonconventional means, so detailed history and open-mindedness continue to be paramount.

HONEYBEE ALLERGENIC COMPONENTS

Honeybee-predominant concerns of VIT efficacy/safety and double-positivity drive continued intense interest in further characterizing HBV components. Table 1 [18–25,26[■],27[■]] lists the current, known HBV allergens. Phospholipase A₂ (Api m 1), a glycoprotein, is the most potent and allergenic protein. Aside from eliciting a response through direct stimulation by cross-linking IgE on mast cells and basophils, it also increases leukotriene production through the hydrolysis of phospholipids [28].

Hyaluronidase (Api m 2) is traditionally thought to be the second most allergenic substance and is found to have about 50% sequence identity with its Vespid counterpart [29]. Recent findings regarding the antigenicity of other HBV allergens, discussed below, seem to challenge the role of hyaluronidase as a major honeybee allergen, especially in light of a recent study [30] showing that cross-reacting carbohydrate determinates (CCDs) account for most of the specific IgE in Vespid hyaluronidase and that much of the cross-reactivity with the honeybee hyaluronidase is likely because of these CCDs.

Melittin (Api m 4) is the largest contributor to HBV composition. It is responsible for much of the pain and inflammation during the sting, likely via cell lysis (red blood cells, myocytes, leukocytes, mast cells, among others) [31]. Although it seems to be a relatively weak allergen, it remains a very important HBV protein due to its abundance, ability to potentiate other venom proteins, and toxicity potential. Api m 3, 5, 6, and 7 show a substantial preponderance for causing sensitization in individuals, with

Table 1. Honeybee venom allergenic components (www.allergen.org), ? = unknown

	Allergen	Size (kD)	Venom (%)	IgE positive (%)	Allergenicity	Glycosylation	Cross-reactivity
Api m 1	Phospholipase A ₂	16	7–15	95 [18]	Very high	Yes	Low
Api m 2	Hyaluronidase	39	1–3	50 [19,20]	Weak to moderate	Yes	High
Api m 3	Acid phosphatase	43	1	37 [21]	Weak	Yes	Moderate
Api m 4	Melittin	3	35–50	28–50 [22]	Weak	No	Low
Api m 5	Dipeptidylpeptidase IV	100	1	60 [23]	?	Yes	Moderate
Api m 6	Cysteine-rich trypsin inhibitor	8	1–2	42 [24]	Weak	No	?
Api m 7	CUB serine protease	39	<1	80 [25]	?	Yes	?
Api m 8	Carboxyesterase	70	<1	?	?	?	?
Api m 9	Serine carboxyesterase	60	?	?	?	?	?
Api m 10	Icarapin	50–55	?	50 [26 [■]]	Moderate to high	Yes	Low
Api m 11	MRJP	–	–	–	–	–	–
11.0101	MRJP 8	50–60	?	15 [27 [■]]	Weak	Yes	?
11.0201	MRJP 9	50–60	?	34 [27 [■]]	Weak	Yes	?
Api m 12	Vitellogenin	200	?	?	?	?	?

CUB, Complement subcomponents C1r/C1s, Uegf, and Bone morphogenic protein-1; MRJP, major royal jelly protein.

reported 30–80% of honeybee-allergic patients being sensitized; however, it is yet unclear in these how much of the sensitivity is owing to CCDs or what is their clinical significance [23–25,32]. Further studies are needed for these potentially allergenically important components.

A fairly recently discovered HBV allergen, icarapin (Api m 10), a carbohydrate-rich protein of unknown function, seems to show genuine and perhaps honeybee-specific allergenicity, exhibiting 50% sensitization among honeybee-allergic patients, regardless of glycosylation (CCDs), which more strongly supports a true sensitivity [26[■]]. This protein, though, seems to be particularly labile, which has made it elusive to analyze, so its true contribution to HBV composition remains unknown. Because of its labile nature, therapeutic immunotherapy may be affected by lack of this substance, as discussed later.

Major royal jelly proteins (Api m 11) have also shown promise as new potential allergens within the HBV. A recent study [27[■]] showed that more than half of patients showed specific IgE to two of these proteins. However, CCDs seemed to contribute a substantial amount to these sensitivities. Specific IgE to the nonglycosylated forms, though, demonstrate that the allergic potential of these proteins should not be ignored.

Another new allergen, vitellogenin (Api m 12), a glycolipoprotein involved in fat storage and deposition, is a protein found in many animals (most notably fish) and their yolks, and it is even an allergen in Vespidae venom (Ves v 6) [33]. Because

of its abundance in many species, this allergen may also account for some degree of cross-reactivity between Hymenoptera venom [34]. It is likely similar to CCDs, being responsible for clinically nonimportant sensitization.

DIFFICULTIES WITH HONEYBEE ALLERGY DIAGNOSIS

The conundrum every allergist faces with venom allergy, particularly with HBV allergy, is the double-positive or multipositive patient, especially with a history of a single sting or multiple stings to a single, known insect. Double-positivity may be because of either true double-positive allergy or cross-reacting substances such as shared amino-acid sequences between venoms or because of CCDs. Methods to clarify these difficult patients have been recently researched and hotly debated.

Current diagnostic standard

According to current guidelines in the United States, evaluation of a patient with a concerning history begins with skin testing using extracted honeybee venom standardized to Api m 1 [35]. Intradermal doses start typically at a concentration of 0.001 µg/ml and progress to 1 µg/ml. These concentrations produce sensitivity, based on strong clinical history, of more than 95% [36,37]. The specificity, however, of intradermal testing may be considerably lower (70–75%), likely because of the myriad of disruptive substances in HBV.

Current serologic methods

Current methods for serologic testing, using immunoassay capture testing (CAP), a solid-phase immunoassay, to determine honeybee venom sensitization use whole venom, again, standardized to Api m 1. The sensitivity and specificity of current methods, based on strong clinical history, is about 93% and 84%, at best, respectively [38]. Though most feel serologic testing is second-line, some allergists may consider using CAP-specific IgE even as their primary test, which is not unreasonable.

Recombinant allergen serologic testing

As it continues to advance, serologic testing seems to be the way of the future for HBV allergy diagnostic testing for several reasons: convenience to the patient (less time, less pain, ability to stay on antihistamine medication), safety (no chance of reaction compared with at least a minimal chance to react to the skin testing), and soon a more refined detection of venom allergen sensitivity, especially with the advent of recombinant allergen technology utilizing component-resolved diagnostics (CRD). Several studies [39–41,42^{***}], using recombinant Api m 1 (rApi m 1) as a surrogate for honeybee allergy detection via serologic CAP, show a sensitivity and specificity, based on clinical history and skin testing, of 60–80% and 100%, respectively. By comparison, Vespid venom components (using both rVes v 1 and rVes v 5) showed a sensitivity of about 94% [43]. Several of these studies concluded as well that many variables, such as geographical, methodological, material, and population differences, may have accounted for the wide range of sensitivities.

Though much correspondence debated the usefulness of rApi m 1 in the diagnosis of HBV allergy, all universally agreed that the sensitivity could be dramatically improved by adding several other recombinant HBV allergens; however, this has not been specifically well studied yet. Although CRD may be good enough one day to be the primary diagnostic tool, its reasonable sensitivity and excellent specificity may make it good enough now as a limited tool to help distinguish double-positivity, as has been proposed [42^{***}]. Unfortunately, CRD for HBV is not currently available for use in the United States, though rApi m 1 is available for commercial use in Europe. As it develops, cost, reproducible and reliable results among laboratories, and access of the testing to allergists are all concerns and potential drawbacks to address.

Role of cross-reacting carbohydrate determinants

CCDs remain at the forefront of discussion and were initially confirmed by one group to elicit more

double-positivity in honeybee-allergic patients [44]. Recently, another group confirmed this and showed that the vast majority (81%) of double-positive, anti-CCD IgE-positive patients were positive to rApi m 1 versus 64% to rVes v 5 [40]. By looking at their interesting data, it can also be concluded that roughly 25% of the double-positive patients can be attributed to CCDs, three-eighths to true double-positivity to rApi m 1 and rVes v 5, and three-eighths it seems to other factors, which may be specific IgE to other venom components (e.g. Api m 3, Api m 10, among others), cross-reacting components (e.g. Api m 2 and Ves v 2), or yet unrecognized allergens. It may have been of interest to stratify the double-positive/CCD data according to specific insect allergy. This may offer further insight into what role the anti-CCD IgE may truly play in honeybee allergy, in particular.

Use of purified honeybee allergens in diagnosis

Purified, natural Api m 1 (nApi m 1) has been studied recently and postulated to help resolve double-positivity. One group concluded that although nApi m 1 was more sensitive, rApi m 1 seemed to catch more truly sensitized individuals, owing to its absence of CCDs [41]. This conclusion was countered by data from another group showing that nApi m 1 was more positive than rApi m 1 in patients with more severe reactions, suggesting that using solely rApi m 1 would miss some critical patients [45]. What cannot be concluded from either of these studies is whether there are simply other substances that have greater clinical significance, such as specific IgE to other HBV allergens, and that the greater sensitivity of nApi m 1 is simply getting 'lucky' by inadvertently being positive because of anti-CCD IgE that is lacking on rApi m 1. What can be concluded is that we are far from understanding double-positive patients, yet a little closer at the same time, and perhaps by continuing studies to include more recombinant HBV components, we will gain further insight.

Basophil activation testing and other considerations

The basophil activation test (BAT) continues to be revisited as a means to distinguish double-positivity. One group recently showed that BAT may compare well against recombinant allergen-based IgE to honeybee and Vespid when evaluating the double-positive patient and may even prove to be a 'next-step' test to distinguish those who are missed by standard methods and/or CRD [46]. The BAT

procedure, however, is wrought with challenges such as procedures varying among laboratories, end points varying among studies reported (i.e. 15% basophil activation versus 50% or greater as a cut off), and being technically challenging and time consuming for the laboratory technician. Specific IgE-inhibition testing can be useful in distinguishing double-positivity as well; however, like BAT, and perhaps more so, it remains technically difficult and time-intensive and is typically only performed in the research setting.

SAFETY AND EFFICACY OF HONEYBEE IMMUNOTHERAPY

Immunotherapy remains the main treatment for preventing future sting reactions by providing long-lasting protection [35]. Compared with Vespid VIT, honeybee VIT is less effective and appears to elicit more systemic reactions, making it less well tolerated as well. For these reasons, studies of late are focused on these areas. A recent, comprehensive review specifically on honeybee VIT discussed many of these findings among other important considerations and is worth the reader's review [47[¶]]. The mechanism of VIT to honeybee and Vespid was also recently thoroughly reviewed [48].

Evaluating venom immunotherapy efficacy

No recent study has evaluated methods to exclusively improve the efficacy of honeybee VIT. However, some studies, discussed below, designed mainly to improve safety, also demonstrated non-inferior effective treatment. Because of the poorer efficacy of honeybee VIT, one study [49[¶]] sought to find a more consistent way to evaluate for VIT effectiveness in a patient by using a microsyringe injection of natural HBV as a surrogate for a real honeybee sting. The syringe method was at least as effective as a natural honeybee sting. This concept of using a surrogate for a true honeybee sting is new, and may show promise for standardizing a type of 'sting challenge' by syringe with known components, thus eliminating the inconsistency of honeybee sting challenges. This may hold diagnostic implications as well.

Another concept, introduced earlier in this article, considers that honeybee VIT may be less efficacious because of the absence of specific HBV allergens, perhaps lost during processing of the venom extract. Api m 10 was not found in any measureable concentration in therapeutic HBV preparations, though it appears to be a significant allergen [26^{¶¶}]. Though there seem to be few data,

lack of specific allergens in HBV extract may explain, at least in part, lower honeybee VIT efficacy, and future research should be strongly considered in this area.

Overall venom immunotherapy safety

A recently published Cochrane review affirmed, through thorough search and strict inclusion criteria, that honeybee VIT is less well tolerated with a systemic reaction rate of 14.2 versus 2.8% for Vespid VIT [50]. Another all-inclusive systematic review of the safety of VIT showed a higher systemic reaction rate for the honeybee VIT of about 25% [51[¶]]. Both of these reviews confirm well established data regarding a less-than-optimal safety profile for honeybee VIT.

Safety of aqueous versus depot extracts

In an effort to improve honeybee VIT safety, the review group from above attempted to separate aqueous extracts from depot extracts. Although, there was a difference in systemic reaction rates between depot and aqueous injections for honeybee VIT, the authors pointed out that this difference may be attributed to rush and cluster protocols for the aqueous and depot injections, respectively [51[¶]]. In another recent study [52^{¶¶}], the authors felt, through cutting-edge spectrometry techniques, that tissue injected with aqueous extract showed greater amounts of dopamine, histidine, norepinephrine, and leukotrienes, suggesting an increased risk for adverse reactions. They also showed that HBV allergens remained in tissue longer following depot injections. Little can be concluded from these studies clinically, but certainly this testing confirms the prolongation of depot preparations and suggests reasons for their improved safety. This study also opens the door for further tissue pharmacokinetic studies that can be of use for the design of future effective HBV extracts and evaluation of specific HBV components.

Safety of purified versus nonpurified extracts

Purified HBV is devoid of the low-molecular-weight proteins that may be responsible, at least in part, for some of the adverse events in honeybee allergy and VIT. One group prospectively found a significant reduction in the systemic reaction rate of 2.5 versus 27.5% in the purified versus nonpurified groups, respectively [53[¶]]. Another group also showed similar low systemic reaction rates [54]. Interestingly, this latter group found that regardless of build-up schedule, the systemic reaction rates varied

little. Although both studies had some weaknesses (low numbers, nonblinded, nonrandomized), these are the first comparative, prospective studies evaluating what appears to be a well tolerated and likely equally effective alternative to the current honeybee VIT standard.

Tryptase and honeybee venom immunotherapy

For those on VIT, use of HBV is an independent risk factor for a systemic reaction, but interestingly, an elevated tryptase did not seem to put those on honeybee VIT at any higher risk, unlike for those on Vespid VIT [55]. A more recent study [56] using decreasing BAT positivity as a measure of protection also demonstrated that honeybee VIT systemic reaction rate did not correlate with tryptase levels. For now, mild tryptase elevation does not seem to be a factor for honeybee allergic patients. Caution should be taken, however, to not misinterpret this to mean that those with mast cell disorders are at no higher risk, as this group represents a separate entity entirely, recently discussed thoroughly elsewhere [57].

Basophil activation testing

One group has recently shown in two separate studies [56,58] that decreasing BAT positivity seems to correlate with honeybee VIT efficacy in children. Though this may one day be more mainstream, as previously discussed, concerns regarding BAT still make this testing method less useful to the practicing allergist, but certainly a considered option.

CONCLUSION

Allergy to honeybees poses unique problems for the allergist. Their complex venom composition, lack of more thorough understanding of HBV allergens, and the enigma of the double-positive honeybee-allergic patients lead to difficulty in diagnosis and management. Fortunately, CRD, recombinant technology, and clever allergists appear to be leading us to a greater understanding of effective solutions.

Acknowledgements

The authors specially thank Scott Dickson, D.O., for use of a fraction of his data.

Conflicts of interest

There are no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

Additional references related to this topic can also be found in the Current World Literature section in this issue (p. 454).

1. Almeida RA, Olivo TE, Mendes RP, *et al.* Africanized honeybee stings: how to treat them. *Rev Soc Bras Med Trop* 2011; 44:755–761.
2. Vetter RS, Visscher PK, Camazine S. Mass envenomations by honey bees and wasps. *West J Med* 1999; 170:223–227.
3. Onbasi K, Onbasi O, Eminbeyli L, Kaynak C. Prevalence and alternative therapy methods for bee and wasp allergy in Van. *Allergy* 2008; 63:246–247.
4. Charpin D, Birnbaum J, Lanteaume A, Vervloet D. Prevalence of allergy to Hymenoptera stings in different samples of the general population. *J Allergy Clin Immunol* 1992; 90:331–334.
5. Incorvaia C, Mauro M, Pastorello EA. Hymenoptera stings in conscripts. *Allergy* 1997; 52:680–681.
6. Barnard JH. Studies of 400 Hymenoptera sting deaths in the United States. *J Allergy Clin Immunol* 1973; 52:259–264.
7. Green A, Breisch NL. Avoidance of bee and wasp stings: an entomological perspective. *Curr Opin Allergy Clin Immunol* 2005; 5:337–341.
8. Breed MD, Guzman-Novoa E, Hunt GJ. Defensive behavior of honey bees: organization, genetics, and comparisons with other bees. *Annu Rev Entomol* 2004; 49:271–298.
9. Ono M, Terabe H, Hori H, Sasaki M. Components of giant hornet *aleram* pheromone. *Nature* 2003; 424:637–638.
10. Woyke J. Diurnal and seasonal variation in defensive behavior of African bees *Apis mellifera adansonii* in Ghana. *Apidologie* 1992; 23:311–322.
11. Brown TC, Tankersley MS. Sting of the honeybee: an allergic perspective. ■ *Ann Allergy Asthma Immunol* 2011; 107:463–470.
12. This is a complete overview of unique aspects of the honeybee insect and honeybee allergy and an overall review of honeybee allergy literature.
13. Hoffman DR, Jacobson RS. Allergens in Hymenoptera venom XII. How much protein is in a sting? *Ann Allergy* 1984; 52:276–278.
14. Schumacher MJ, Tveten MS, Egen NB. Rate and quantity of delivery of venom from honeybee stings. *J Allergy Clin Immunol* 1994; 93:831–835.
15. Hsu CY, Chiang WC, Weng TI, *et al.* Laryngeal edema and anaphylactic shock after topical propolis use for acute pharyngitis. *Am J Emerg Med* 2004; 22:432–433.
16. Katayama M, Aoki M, Kawana S. Case of anaphylaxis caused by ingestion of royal jelly. *J Dermatol* 2008; 35:222–224.
17. Dickson S. MC contact with HB venom abstract-Harold S Nelson military symposium-Mar 2012 [Online]. Email to Tyson Brown (tyson.brown@us.af.mil) 2012 Feb 23 [cited 2012 March 2].
18. Shin YS, Liu JN, Hur G-Y, *et al.* Clinical features and the diagnostic value of component allergen-specific IgE in Hymenoptera venom allergy. *Allergy Asthma Immunol Res* 2012; 4:284–289.
19. Dudler T, Scheider T, Annand RR, *et al.* Antigenic surface of bee venom phospholipase A2. *J Immunol* 1994; 152:5514–5522.
20. Soldatova LN, Cramer R, Gmachl M, *et al.* Superior biologic activity of the recombinant bee venom allergen hyaluronidase expressed in baculovirus-infected insect cells as compared with *Escherichia coli*. *J Allergy Clin Immunol* 1998; 101:691–698.
21. Markovic-Housley Z, Miglierini G, Soldatova L, *et al.* Crystal structure of hyaluronidase, a major allergen of bee venom. *Structure* 2000; 8:1025–1035.
22. de Graaf DC, Aerts M, Danneels E, Devreese B. Bee, wasp, and ant venomomics pave the way for a component-resolved diagnosis of sting allergy. *J Proteomics* 2009; 72:145–154.
23. Kemeny DM, Harries MG, Youlten LJ, *et al.* Antibodies to purified bee venom proteins and peptides. I. Development of a highly specific RAST for bee venom antigens and its application to bee sting allergy. *J Allergy Clin Immunol* 1983; 71:505–514.
24. Blank S, Seismann H, Bockisch B, *et al.* Identification, recombinant expression and characterization of the 100 kDa high molecular weight Hymenoptera venom allergens Api m5 and Ves v3. *J Immunol* 2010; 184:5403–5413.
25. Kettner A, Hughes GJ, Frutiger S, *et al.* Api m6: A new bee venom allergen. *J Allergy Clin Immunol* 2001; 107:914–920.
26. Winningham KM, Fitch CD, Schmidt M, Hoffman DR. Hymenoptera venom protease allergens. *J Allergy Clin Immunol* 2004; 114:928–933.
27. Blank S, Seismann H, Michel Y, *et al.* Api m 10, a genuine *A. mellifera* venom ■ allergen, is clinically relevant but underrepresented in therapeutic extracts. *Allergy* 2011; 66:1322–1329.
28. This is the first study demonstrating the ability to produce and evaluate a previously elusive honeybee allergen, icarapin, Api m 10. The authors also showed that this protein may be a major honeybee venom allergen, and current immunotherapy extracts may not afford protection, given that no traceable amounts of the protein were found during analysis of the extracts.

27. Blank S, Bantleon FI, McIntyre M, *et al.* The major royal jelly proteins 8 and 9 (Apm 11) are glycosylated components of *Apis mellifera* venom with allergenic potential beyond carbohydrate-based reactivity. *Clin Exp Allergy* 2012; 42:976–985.

This is the first study cloning two new, related honeybee proteins. The authors also demonstrated that these proteins appear to be allergenic, though affected by cross-reacting carbohydrate determinants.

28. Mustafa FB, Ng FS, Nguyen TH, Lim LH. Honeybee venom secretory phospholipase A2 induces leukotriene production but not histamine release from human basophils. *Clin Exp Immunol* 2008; 151:94–100.
29. King TP, Spangfort MD. Structure and biology of stinging insect venom allergens. *Int Arch Allergy Immunol* 2000; 123:99–106.
30. Jin C, Focke M, Leonard R, *et al.* Reassessing the role of hyaluronidase in yellow jacket venom allergy. *J Allergy Clin Immunol* 2010; 125:184–190.
31. Hoffman DR. Hymenoptera venoms: composition, standardization, stability. In: Levine MI, Lockey RF, editors. *Monograph on insect allergy*, 4th Ed. Pittsburgh: Dave Lambert Associates; 2003. pp. 37–53.
32. Grunwald T, Bockisch B, Spillner E, *et al.* Molecular cloning and expression in insect cells of honey bee venom allergen acid phosphatase (Api m3). *J Allergy Clin Immunol* 2006; 117:848–854.
33. Allergen Nomenclature [Internet]. [place unknown]: World Health Organization/International Union of Immunological Societies Allergen Nomenclature Sub-committee; [date unknown] [cited 2013 Mar 25]. <http://www.allergen.org>.
34. Piulachs MD, Guidugli KR, Barchuk AR, *et al.* The vitellogenin of the honeybee, *Apis mellifera*: structural analysis of the cDNA and expression studies. *Insect Biochem Mol Biol* 2003; 33:459–465.
35. Golden DBK, Moffitt J, Nicklas RA. Stinging insect hypersensitivity: a practice parameter update 2011. *J Allergy Clin Immunol* 2011; 127:852–854.
36. Patrizzi R, Müller U, Yman L, Hoigne R. Comparison of skin tests and RAST for the diagnosis of bee sting allergy. *Allergy* 1978; 34:249–256.
37. Day JH, Buckeridge DL, Welsh AC. Risk assessment in determining systemic reactivity to honeybee stings in sting-threatened individuals. *J Allergy Clin Immunol* 1994; 93:691–705.
38. Scherer K, Weber JM, Jermann TM, *et al.* Cellular in vitro assays in the diagnosis of Hymenoptera venom allergy. *Int Arch Allergy Immunol* 2008; 146:122–132.
39. Sturm GJ, Hemmer W, Hawranek T, *et al.* Detection of IgE to recombinant Api m 1 and r Ves v 5 is valuable but not sufficient to distinguish bee from wasp venom allergy. *J Allergy Clin Immunol* 2011; 128:247–248.
40. Hofmann SC, Pfender N, Weckesser S, *et al.* Added value of IgE detection to rApi m 1 and rVes v 5 in patients with Hymenoptera venom allergy. *J Allergic Clin Immunol* 2011; 127:265–267.
41. Jakob T, Köhler J, Blank S, *et al.* Comparable IgE reactivity to natural and recombinant Api m 1 in cross-reactive carbohydrate determinant-negative patients with bee venom allergy. *J Allergy Clin Immunol* 2012; 130:276–278.
42. Müller U, Schmid-Grendelmeier P, Hausmann O, Helbling A. IgE to recombinant allergens Api m 1, Ves v 1, and Ves v 5 distinguish double sensitization from crossreaction in venom allergy. *Allergy* 2012; 67:1069–1073.
- Though this is not the first study evaluating the sensitivity of using recombinant allergens, these authors suggest a very reasonable approach to using skin testing, serologic recombinant allergens, and even basophil activation testing when presented with the difficult double-positive patient.
43. Ebo DG, Faber M, Sabato V, *et al.* Component-resolved diagnostics of wasp (yellow jacket) venom allergy. *Clin Exp Allergy* 2012; 43:255–261.
44. Müller UR, Johansen N, Petersen AB, *et al.* Hymenoptera venom allergy: analysis of double positivity to honey bee and *Vespula* venom by estimation of IgE antibodies to species-specific major allergens Api m1 and Ves v5. *Allergy* 2009; 64:543–548.

45. Korošec P, Valenta R, Mittermann I, *et al.* Low sensitivity of commercially available rApi m 1 for diagnosis of honeybee venom allergy. *J Allergy Clin Immunol* 2011; 128:671–673.

46. Eberlein B, Krischan L, Darsow U, *et al.* Double positivity to bee and wasp venom: improved diagnostic procedure by recombinant allergen-based IgE testing and basophil activation test including data about cross-reactive carbohydrate determinants. *J Allergy Clin Immunol* 2012; 130:155–161.

47. Bilò MB, Antonicelli L, Bonifazi F. Honeybee venom immunotherapy: ■ certainties and pitfalls. *Immunotherapy* 2012; 4:1153–1166.

This is a very comprehensive, thorough, and up-to-date review of HBV immunotherapy that covers nearly every aspect.

48. Özdemir C, Kucuksezer UC, Akdis M, Akdis CA. Mechanisms of immunotherapy to wasp and bee venom. *Clin Exp Allergy* 2011; 41:1226–1234.

49. Cortellini G, Severino M, Francescato E, *et al.* Evaluation and validation of a ■ bee venom sting challenge performed by a micro-syringe. *Ann Allergy Asthma Immunol* 2012; 109:438–441.

In an effort to create a consistent tool for evaluation of immunotherapy efficacy, these authors showed a preliminary study that may pave the way for a reproducible and perhaps ethical sting challenge via microsyringe.

50. Boyle RJ, Elremeli M, Hockenhull J, *et al.* Venom immunotherapy for preventing allergic reactions to insect stings. *Cochrane Database Syst Rev* 2012; (10):CD008838.

51. Incorvaia C, Frati F, Dell'Albani I, *et al.* Safety of Hymenoptera venom ■ immunotherapy: a systematic review. *Expert Opin Pharmacother* 2011; 12:2527–2532.

Though not new data, this is the most complete summarization of all honeybee and Vespid safety data for the last 25 years, with some unique insight into some of the findings.

52. Seppälä U, Francese S, Turillazzi S, *et al.* In situ imaging of honeybee (*Apis ■ mellifera*) venom components from aqueous and aluminum hydroxide-adsorbed venom immunotherapy preparations. *J Allergy Clin Immunol* 2012; 129:1314–1320.

This is the first study to use a cutting-edge imaging tool to look at the absorption of various HBV components in live tissue over time. The findings confirm that depot shots indeed have a depot effect for the allergens and suggest an explanation of why aqueous injections may produce more reactions. This may be the first of many similar studies using this imaging tool, as we seem to be at the tip of its application.

53. Bilò MB, Cinti B, Brianzoni MF, *et al.* Honeybee venom immunotherapy: a ■ comparative study using purified and nonpurified aqueous extracts in patients with normal basal serum tryptase concentrations. *J Allergy (Cairo)* 2012; 2012:869243.

This is the first study prospectively comparing the systemic reaction rates of purified versus nonpurified extracts in immunotherapy.

54. Patella V, Florio G, Giuliano A, *et al.* Hymenoptera venom immunotherapy: tolerance and efficacy of an ultrarush protocol versus a rush and a slow conventional protocol. *J Allergy (Cairo)* 2012; 2012:192192.

55. Ruëff F, Przybilla B, Bilò MB, *et al.* Predictors of side effects during the buildup phase of venom immunotherapy for Hymenoptera venom allergy: the importance of baseline serum tryptase. *J Allergy Clin Immunol* 2010; 126:105–111.

56. Žitnik SEK, Vesel T, Avčin T, *et al.* Monitoring honeybee venom immunotherapy in children with the basophil activation test. *Pediatr Allergy Immunol* 2012; 23:166–172.

57. Gonzalez de Olano D, Alvares-Twose I, Vega A, *et al.* Venom immunotherapy in patients with mastocytosis and Hymenoptera venom anaphylaxis. *Immunotherapy* 2011; 3:637–651.

58. Eržen R, Košnik M, Šilar M, Korošec P. Basophil response and the induction of a tolerance in venom immunotherapy: a long-term sting challenge study. *Allergy* 2012; 67:822–830.